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## **The evolutionary puzzle of egg size, oxygenation and parental care in aquatic environments**

Braga Goncalves, Ines ; Ahnesjö, Ingrid ; Kvarnemo, Charlotta

**Abstract:** Offspring fitness generally improves with increasing egg size. Yet, eggs of most aquatic organisms are small. A common but largely untested assumption is that larger embryos require more oxygen than they can acquire through diffusion via the egg surface, constraining egg size evolution. However, we found no detrimental effects of large egg size on embryo growth and survival under hypoxic conditions. We tested this in the broad-nosed pipefish, *Syngnathus typhle*, whose males provide extensive care (nourishment, osmoregulation and oxygenation) to their young in a brood pouch on their bodies. We took advantage of this species' pronounced variation in egg size, correlating positively with female size, and tested the effect of hypoxia (40% dissolved oxygen) versus fully oxygenated (100%) water on embryo size and survival of large versus small eggs after 18 days of paternal brooding. Egg size did not affect embryo survival, regardless of O<sub>2</sub> treatment. While hypoxia affected embryo size negatively, both large and small eggs showed similar reductions in growth. Males in hypoxia ventilated more and males with large eggs swam more, but neither treatment affected their position in the water column. Overall, our results call into question the most common explanation for constrained egg size evolution in aquatic environments.

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**The evolutionary puzzle of egg size, oxygenation and parental care in aquatic environments**

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1                   **THE EVOLUTIONARY PUZZLE OF EGG SIZE, OXYGENATION AND**  
2                   **PARENTAL CARE IN AQUATIC ENVIRONMENTS**

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**Abstract**

Offspring fitness generally improves with increasing egg size. Yet, eggs of most aquatic organisms are small. A common but largely untested assumption is that larger embryos require more oxygen than they can acquire through diffusion via the egg surface, constraining egg size evolution. However, we found no detrimental effects of large egg size on embryo growth and survival under hypoxic conditions. We tested this in the broad-nosed pipefish, *Syngnathus typhle*, whose males provide extensive care (nourishment, osmoregulation, oxygenation) to their young in a brood pouch on their bodies. We took advantage of this species' pronounced variation in egg size, correlating positively with female size, and tested the effect of hypoxia (40% dissolved oxygen) vs. fully-oxygenated (100%) water on embryo size and survival of large vs. small eggs after 18 days of paternal brooding. Egg size did not affect embryo survival, regardless of O<sub>2</sub> treatment. Whilst hypoxia affected embryo size negatively, both large and small eggs showed similar reductions in growth. Males in hypoxia ventilated more and males with large eggs swam more, but neither treatment affected their position in the water column. Overall, our results call into question the most common explanation for constrained egg size evolution in aquatic environments.

43 **INTRODUCTION**

44 Large juveniles commonly enjoy fitness benefits compared to smaller ones [1] and juvenile  
 45 and adult size are often positively related to initial egg size [2]. Should we therefore predict  
 46 evolution of ever increasing egg sizes? Following Smith and Fretwell's [3] influential model,  
 47 we should not due to the trade-off between offspring size and number. Moreover, even though  
 48 larger offspring show better survival and/or fitness, they do so with diminishing returns [3].  
 49 However, this is not the only cost or trade-off associated with increasing egg size in aquatic  
 50 environments. Because there is much lower O<sub>2</sub> availability in water than in air, oxygenation  
 51 can be a challenging process in aquatic environments and has commonly been argued to  
 52 constrain egg size evolution [4-6]. Indeed, most fish eggs are spherical, the shape most  
 53 favourable for gas exchange in low water velocities [7]. However, given a round shape, an  
 54 increase in egg size results in a greater increase of volume compared to the surface area.  
 55 Hence, the relative surface area, where diffusion takes place, decreases as volume increases  
 56 [4, 8]. Assuming that egg volume determines embryonic oxygen (O<sub>2</sub>) requirements and egg  
 57 surface area determines the ability to acquire O<sub>2</sub> by diffusion, limitations in O<sub>2</sub> availability to  
 58 the embryos may constrain egg size evolution. Yet, whilst O<sub>2</sub> limitations have been amply  
 59 demonstrated in egg masses [9-11] they have only been assumed to affect individual embryos  
 60 in response to egg size. In fact, despite theoretical support for the principle [reviewed in 8], a  
 61 negative effect of egg size on embryo survival due to limited O<sub>2</sub> uptake is yet to be confirmed  
 62 [4, 6, 12, 13].

63

64 The amount of dissolved O<sub>2</sub> in water affects the metabolism and development of embryos [8,  
 65 14, 15] and consequently their survival. A decrease in dissolved oxygen (DO) concentrations  
 66 in the water can lead to hypoxia [i.e. any level of reduced DO that negatively affects the  
 67 physiology or behaviour of an organism, 16], which can reduce embryo metabolism [17],

decrease yolk conversion efficiency [18], reduce development rates and delay hatching times [19], as well as induce premature hatching [19], cause deformities [20], decrease size at hatching and reduce post-hatching survival [19]. The hypothesis that large eggs do worse than smaller ones assumes that  $O_2$  consumption increases proportionally with egg volume [4-6]. However, if  $O_2$  consumption increases at a slower rate than volume, smaller eggs can be predicted to perform worse [4]. This was demonstrated in Atlantic salmon, *Salmo salar*, where embryo  $O_2$  requirements do not increase in direct proportion to the volume, suggesting that at low DO concentrations, embryos of small eggs, and not of large ones, may have difficulties obtaining sufficient  $O_2$  for their development [4].

In a development of Smith and Fretwells' (1974) model, Jorgensen et al. (2011) showed that optimal offspring size generally decreases with increasing growth rate. However, when the authors included parental care in the model in the form of live bearing, offspring survival became a function of parental survival, with larger parents predicted to produce larger offspring. This prediction is in accordance with interspecific patterns among ectothermic animals, which indicate that parental care and egg size are positively associated [21-23]. In aquatic environments, how and whether parental care and egg size coevolved is complex to disentangle, but parental care often involves the task of oxygenation of developing embryos [6, 23]. In giant water bugs (Belostomatidae) selection on egg size and embryo oxygenation are key factors in the evolution of paternal care [24]. Males carry the relatively large eggs on their backs and promote  $O_2$  diffusion through the egg membranes by "pumping" the brood in and out of water or by brushing the eggs with their hind legs [24]. Among fishes there is substantial variation in egg size [25, 26], with larger egg sizes found in demersally than in pelagically spawning species [25]. Concurrently, parental care is more common in the former than in the latter group [27] and a variety of parental care strategies directly improve the

93 access of O<sub>2</sub> to the embryos. For example, parents may choose nest sites according to O<sub>2</sub>  
 94 availability [28, 29], fan their eggs to improve suboptimal O<sub>2</sub> conditions [30, 31], increase  
 95 nest opening size [32, 33] or cannibalise some of the eggs to improve the oxygenation of the  
 96 remaining embryos [34, 35]. In fact, parental care in aquatic species may have evolved, at  
 97 least partially, to protect developing embryos from low O<sub>2</sub> availability, which in turn could  
 98 have favoured the evolution of larger egg sizes [6, 36].

99

100 The aim of the current study was to investigate if egg size and O<sub>2</sub> availability interact as  
 101 expected based on the hypothesis that egg size is evolutionarily constrained by O<sub>2</sub>-acquiring  
 102 ability in aquatic environments. We tested this by examining embryo development and  
 103 survival in the broad-nosed pipefish, *Syngnathus typhle* L., a species with substantial paternal  
 104 care and pronounced variation in egg size, with large females producing eggs significantly  
 105 larger than those of small females [37]. Together with the fact that it provides extensive  
 106 parental care and lives in an environment in which ambient O<sub>2</sub> levels vary naturally from  
 107 hypoxic to hyperoxic [38-41], these characteristics make it a very apt model organism. As is  
 108 typical for many pipefishes and seahorses (Syngnathidae), males of this genus care for the  
 109 developing embryos in a sealed brood pouch where they protect, osmoregulate, oxygenate and  
 110 transfer nutrients to the embryos through the vascularized walls of the pouch [37, 42-45].  
 111 Moreover, it is known from previous work on *S. typhle* that larger embryos have higher  
 112 respiration rates than smaller ones [46], larger eggs result in larger offspring [47] and that  
 113 larger juveniles enjoy higher fitness benefits [48].

114

115 We tested whether *S. typhle* embryos are negatively affected by hypoxia during development  
 116 and whether embryos developing from large eggs are more negatively affected by hypoxia  
 117 than embryos developing from small eggs. In addition, we collected behavioural data from the

brooding males to assess whether activity patterns and opercular movements, indicative of ventilation rates, are affected by the size of the eggs they brood or the ambient O<sub>2</sub> saturation levels they experience. The position of the brooding males was also recorded as O<sub>2</sub> saturation levels may vary in the water column. This study provides a valuable contribution to the understanding of how limited access to O<sub>2</sub> affects embryo development and relates to egg size evolution in aquatic environments.

## METHODS AND MATERIALS

### *Fish collection*

This study was conducted at the Swedish West coast at the Sven Lovén Centre for Marine Sciences, Kristineberg (58°15' N, 11°28' E). Due to time and space limitations, the study was done over two consecutive summers (2007 and 2008). *S. typhle* were collected from shallow eelgrass (*Zostera marina*) meadows in the vicinity of the marine station using a beam trawl pulled behind a boat. For details on fish husbandry please see the Supplementary Material.

### *Experimental Design*

Males were randomly allocated to one of four treatment groups: males brooding small or large eggs in either fully oxygenated or in hypoxic water (large eggs, high oxygen (O<sub>2</sub>): n = 12; large eggs, low O<sub>2</sub>: n = 11; small eggs, high O<sub>2</sub>: n = 12; small eggs, low O<sub>2</sub>: n = 12). All males and females had their standard body length (SL) measured, to the nearest mm, before being introduced into the mating tanks. Only medium sized males (mean SL ± SE: 170.3 ± 0.9 mm, range = 155 - 181 mm, n = 47) were used to limit potential size-related differences in male condition, feeding ability, quality of paternal care provided to the embryos and maternal effects resulting from female choice [49]. Male SL did not differ between treatments (1-way ANOVA: F<sub>3,43</sub> = 0.41, p = 0.75). Since female SL is significantly and positively correlated



with egg size [37], females of small (range 160 – 180 mm) and large (230 – 260 mm) sizes were chosen to obtain two distinct egg size categories. Males brooded on average  $85 \pm 25$  eggs (mean  $\pm$  SD, range 25 – 138) which did not differ between O<sub>2</sub> treatments (t-test:  $t = 0.90$ ,  $p = 0.37$ ,  $df = 45$ ). However, males mated with large females received 17% fewer eggs than males mated with small females, and thus, brooding small eggs (t-test:  $t = 2.23$ ,  $p = 0.03$ ,  $df = 45$ , small eggs:  $92 \pm 25$  eggs, large eggs:  $77 \pm 24$  eggs, mean  $\pm$  SD). Using the equation of the relationship between female body length and egg diameter ( $y = 0.0026X + 1.3298$ ) from [48], to calculate egg surface areas for the two egg size treatments showed that surface area of small eggs was on average  $9.86 \text{ mm}^2$ , whereas large eggs were  $12.15 \text{ mm}^2$ , resulting in total egg surface areas of  $907.30 \text{ mm}^2$  for broods of small eggs, and  $935.55 \text{ mm}^2$  for broods of large eggs. Thus, the 17% difference in number of eggs brooded by the males in the two egg size treatments translated into only a 3% difference in egg surface area between treatments.

*S. typhle* is a polygynandrous species, and in our study population, males typically brood eggs from 3-4 females per pregnancy [50-53]. Females produce eggs continuously [54] and large females produce enough eggs to fill up almost three similar-sized males during the course of one pregnancy, whilst small females can fill up little more than one male of similar size during the same time period [55, 56]. Thus, males were allowed to mate polygynously, by introducing groups of males in large mating aquaria (approx. 70 L) containing groups of either large or small females. Specifically, females and males were introduced in the mating tanks in ratios of 1:2 for large females (four females and eight males) and 1:1 for small females (eight females and eight males), and individual females were replaced periodically if they looked egg-limited, to ensure that males mated and filled up their brood pouches within a few days. All matings took place in fully oxygenated water.

Each male was kept in the mating tank until his pouch was filled with eggs. Once fully mated, the male was measured and transferred into a smaller (26 x 45 x 40 cm or 26 x 35 x 35 cm) individual tank, where he was kept brooding for 18 days at a high or low level of O<sub>2</sub> saturation (details below). After 18 days the embryos have developed eyespots and this period corresponds approximately to between one third and one half of the total brooding period at 14-15°C [56].

Brooding aquaria were supplied with either fully oxygenated water (approx. 100% O<sub>2</sub>, which corresponds to 8.70 mg/L or 156 mm Hg O<sub>2</sub> at 15°C) or hypoxic water (40% O<sub>2</sub>, corresponding to 3.50 mg/L or 63 mm Hg O<sub>2</sub> at 15°C), referred to as high and low O<sub>2</sub> below. In natural seagrass habitats, O<sub>2</sub> saturation varies substantially with light and temperature, often between 60-150% [38-40, 57], but 40% is well within the natural range [40]. The hypoxic level of 40% O<sub>2</sub> saturation was chosen based on previous studies on sand gobies [33, 58], collected from the same bay as our pipefish, where 30-40% O<sub>2</sub> was used. These levels elicited behavioural changes without causing mortality.

DO concentration was decreased by pumping nitrogen into the water with the use of a MiniModule 1.7\* 5.5 Membrane Contactor (Liqui-Cel, Celgard, Inc, North Carolina, USA), which removes the O<sub>2</sub> through a counter-current system (i.e. water and nitrogen flow in opposite directions). This unit allowed hypoxic aquaria to have flow-through water like the fully-oxygenated aquaria (see Supplementary Material for details on maintenance of O<sub>2</sub> treatments).

At day 18 of brooding, males were euthanized by immersion in 1ml 2-phenoxyethanol/L seawater) solution for 10 minutes, followed by severing the spinal column immediately

posterior to the opercula. Each male was preserved in 70% ethanol for later dissection. One male was removed from all analyses because its eggs were all unfertilized.

*Embryo survival and size*

Males were dissected to assess relative embryo survival. This was accomplished by calculating the proportion of well-developing embryos out of all eggs in the brood pouch, including unfertilised eggs and substantially underdeveloped embryos.

To obtain estimates of embryo size for each brood at the end of the 18-day brooding period, the average length and weight of embryos were collected as follows: Fifteen embryos, from separate regions of the brood pouch, were removed from each male. The embryos were separated from the egg membrane and the yolk sac. Photographs were taken of five of the embryos using a camera (Leica DFC420 A) attached to a stereo microscope (Leica MZ16 A). The total length (tip of rostrum to tip of tail) of each embryo ( $\pm 0.01\text{mm}$ ) was measured from the photographs of the five embryos using Leica Application Suite, version 2.7.0.RI (Build: 1294). The remaining ten embryos were placed on a Petri dish in a heating cupboard ( $60^{\circ}\text{C}$ ) for one week after which the dried embryos were weighed twice on a Sartorius LE26P microbalance ( $\pm 2\mu\text{g}$ ). The average of these measurements divided by 10 gave the average embryo weight for each male.

*Male behaviour*

On days 1, 9 and 18 of the brooding period ventilation rates and 10-minute video recordings were taken for each male. Ventilation rates were counted during direct observations as number of opercular movements of each individual for 30 seconds. Videos were analysed using JWatcher v1.0. From the video-recordings, proportion of time spent above the mid-line

of the water column and proportion of time spent actively swimming were recorded. Six males were removed from these analyses because we did not have data on all days for these males.

### *Statistical Analysis*

SPSS 22 (IBM SPSS Statistics for Windows, Version 22.0. Armonk, NY: IBM Corp.) and PERMANOVA+ for PRIMER v6 (PRIMER-E, Plymouth, UK) were used to perform all analyses. PERMANOVA+ was chosen because its tests rely on permutations to calculate the distribution of the data, thus avoiding the strict assumptions of parametric tests that are difficult to meet when analysing groups with unequal sample sizes. Permanova has two assumptions: multivariate observations are independent (which they are between males) and identically distributed, i.e., the dispersion clouds are homogenous. To meet the second assumption, some of the data were first transformed, as detailed below.

Because some of the response variables measured were not independent from each other, e.g., embryo length, dry weight and survival, we chose to analyse our data using multivariate statistics to assess overall effects of our treatments and their interactions on our response variables. Only if the multivariate tests returned a significant effect **did** we then analysed the response variables separately. Permanova was used to assess: a) the effects of year (random factor (RF)), egg size treatment (fixed factor (FF): small and large) and O<sub>2</sub> treatment (FF: high and low), on embryo survival, length and dry weight, and b) the effects of year (RF), day (FF, repeated-measure: days 1, 9 and 18), O<sub>2</sub> treatment (FF), egg size treatment (FF) and male ID (RF, nested within year, egg size treatment and O<sub>2</sub> treatment) on ventilation rates, proportion of time spent swimming and proportion of time spent in the upper part of the water column. In (a), embryo survival was arcsine-transformed and embryo length was log-

transformed and in (b), proportion of time spent swimming and proportion of time spent in the upper half of the aquarium were arcsine-square root-transformed. Response variables were normalized (each variable had its mean subtracted and was divided by its standard deviation) in order to achieve a common scale between the variables before producing a resemblance matrix based on Euclidean distances [59, 60] for each multivariate test. As a rule, interaction terms with  $p$  (perm)  $> 0.2$  were sequentially removed from the models, starting with the highest level of interactions, followed by within-level least significance. Due to its central importance to the questions addressed in this study, the “egg size x oxygen” interaction term was kept in the analyses of embryo survival, length and dry weight. Final models are presented in Tables 1 and 2. Whenever the multivariate permutational analyses showed significant terms, permutational ANOVA’s were performed on the separate response variables, using the same reduced models and transformed variables as in the MANOVA’s. The following options were chosen for all analyses: type III sums of squares, fixed effects sum to zero, model: permutation of residuals under a reduced model, number of permutations: 9999.

Since all matings were done prior to the experimental O<sub>2</sub> treatments the effect of O<sub>2</sub> treatment and of egg size treatment on number of eggs received by the males were analysed separately with t-tests. Significance level for all tests, except for the interactions mentioned above, was set at  $p < 0.05$ .

## RESULTS

### *Embryo survival and size*

Embryo survival was on average 75% (range: 18–97%) in all males combined. There was a significant overall variability between years in embryo survival and size (Table 1a). The

multivariate tests showed strong effects of egg size and O<sub>2</sub> treatments but not of their interaction (Table 1a). Post-hoc tests show that these effects were due to low O<sub>2</sub> level having a strong negative effect both on embryo length (Figure 1a) and on embryo weight (Figure 1b), but not on embryo survival (Table 1b). Egg size showed a strong positive effect on embryo weight and tended to affect embryo length positively, but neither egg size nor O<sub>2</sub> treatment affected embryo survival (Table 1b). If large eggs had done worse in low O<sub>2</sub>, significant interactions would be expected between egg size and O<sub>2</sub> level however, no such interactions were found (Table 1a and 1b).

### *Male behaviour*

There was a significant year by day interaction in the overall behaviours, but behaviours did not change significantly with day in the brooding cycle (Table 2a). Both O<sub>2</sub> and egg size treatments affected overall male behaviour significantly (Table 2a). Individuals differed significantly in their behaviour (Table 2a), particularly in their ventilation rates (Table 2b). Ventilation rates were significantly higher in males kept in low O<sub>2</sub> conditions compared to males kept in high O<sub>2</sub> conditions (Figure 2, Table 2b). Males brooding larger eggs swam significantly more than males brooding smaller eggs, but neither egg size nor O<sub>2</sub> level affected the time they spent in the upper part of the aquaria (Table 2b).

## **DISCUSSION**

### *Egg size*

We found no interactions between egg size and O<sub>2</sub> level on embryo survival or growth, despite the large intraspecific variation in egg size observed in this species [37]. This means that embryos from large eggs did not develop significantly worse than embryos from small eggs in hypoxic conditions, as has been predicted from theory [4, 6, 13]. If embryos from

large eggs have lower metabolic rates per egg volume compared to embryos from small eggs, in similarity to the scaling effect observed in brown trout, *Salmo trutta*, [4], such differences in metabolic rates may be large enough to compensate for the less favourable surface area to volume ratio of large eggs. Empirical support for this premise is mixed. For instance, in the mouth brooding cichlid, *Pseudocrenilabrus multicolour victoriae*, females from populations that experience hypoxic conditions year-round produce smaller yet more numerous eggs than females from populations that consistently experience high O<sub>2</sub> saturation levels [15]. However, a complementary laboratory study showed that F1 females from both populations produced larger eggs in hypoxic than in fully-oxygenated water [15]. This result lends support to the study by Einum and colleagues [4], which showed that brown trout embryos from small eggs had lower survival in hypoxic water compared to embryos from larger eggs. Together these studies have generated some controversy as to which egg size is more negatively affected by hypoxia.

In *S. typhle*, embryo respiration increases with increasing embryo dry mass with a slope of 0.44 [61]. Although the relationship between respiration and egg volume is not exactly known, we know that egg diameter and dry mass are strongly and positively correlated [37]. Thus, this relatively low slope (clearly <1) indicates that embryos from larger eggs have lower metabolic rates per egg volume compared to embryos from smaller eggs. If so, such difference in metabolic rates could explain why embryos from the small and large egg treatments were similarly affected by our hypoxic treatment. Thus, our study adds to the list of studies questioning whether embryos from larger eggs are indeed more O<sub>2</sub>-constrained in aquatic environments.

Our males displayed low swimming activity, consistent with the generally inactive and cryptic behaviour of syngnathids and with previous studies showing that pregnant males swim less than females and non-pregnant males [62, 63]. However, males brooding larger eggs swam significantly more than males brooding smaller eggs, though they did not differ in the time spent in the upper part of the aquaria. This result is in line with the increased swimming activity found in other species, shown to increase the amount of water that passes through the gills, improving O<sub>2</sub> uptake [64, 65]. It also resembles the typical parental behaviour of the giant waterbug, in which fathers improve the access of O<sub>2</sub> to the developing embryos [66]. It is thus possible that swimming in *S. typhle* has similar effects, with the movements of the paternal body facilitating O<sub>2</sub> uptake in the gills and this way improving gas exchange to meet the higher O<sub>2</sub> requirements of larger embryos.

Males that brooded large eggs received 17% fewer eggs than males brooding small eggs. Thus, our data provide a possibly unique example, in which males that mate with large females pay a cost by caring for fewer offspring. Since large females are preferred as mates in this species [61], this clearly suggests that the fitness benefits males gain from that preference [e.g. larger offspring that survive better; 48, 67] are large enough to override the number's cost. Moreover, caring for fewer embryos, i.e. having a lower embryo density in the pouch, may allow for better oxygenation. Interestingly, O<sub>2</sub> levels in the bottom section of the pouch tend to be higher compared to other sections [68] and embryo density is often lower in this part of the pouch (personal observation). However, whether egg size affects O<sub>2</sub> levels in the pouch still remains to be tested.

*Oxygenation*



The males brooding in hypoxia showed significantly faster opercular movements throughout the experiment. Faster opercular movements result in faster O<sub>2</sub> extraction from the water, increasing O<sub>2</sub> availability for the males' own metabolic needs and, supposedly, for the developing embryos. In species where parents fan the clutches to ensure a steady access of O<sub>2</sub> to the developing embryos, similar increases in ventilation rates with decreasing O<sub>2</sub> availability have been reported [e.g., 31, 33]. Yet, in our study, embryos brooded in low O<sub>2</sub> conditions were shorter and lighter than embryos brooded in normal O<sub>2</sub> conditions, indicating a clear negative effect of hypoxia on embryo development. A previous study that measured O<sub>2</sub> concentrations in the pouch fluid of brooding *S. typhle* kept in normoxia and hypoxia reported that pouch fluid O<sub>2</sub> concentrations were much lower than those of the water surrounding the males in both treatments, and 2) pouch fluid O<sub>2</sub> concentrations were significantly lower in the males kept in hypoxia compared to the males kept in normoxia [68]. Two important conclusions can be drawn from that study: first, it is noteworthy that pouch brooded embryos naturally develop in much poorer O<sub>2</sub> conditions than those of the ambient water (about 40% lower O<sub>2</sub>); and second, that males appear to have limited capacity to buffer the developing embryos from prolonged environmental hypoxia.

Differences in length and weight of embryos brooded in high and low O<sub>2</sub> conditions can arise from several direct and indirect mechanisms. First, under hypoxia, tissue differentiation rates retard development, so that embryos from males kept in low O<sub>2</sub> conditions may be at earlier stages of development resulting in smaller sizes [69, 70]. Second, development may progress at similar rates but embryos end up smaller due to less efficient anaerobic metabolic processes [71, 72]. Thirdly, quality of parental care (for instance, nutrients provided) may differ in normal and low O<sub>2</sub> conditions due to the physiological stress imposed by hypoxia on the caring parent, affecting the developing young indirectly. Both developmental retardation and

smaller sizes at emergence under hypoxia have been recorded in a copepod [*Acartia tonsa*, 73] showing that these mechanisms are not mutually exclusive.

Whether due to anthropogenic effects or natural causes, hypoxia affects aquatic environments worldwide and in particular areas that are important for reproduction to many aquatic organisms, such as estuaries, semi-enclosed areas and shallow coastal regions [74-76]. In this study, we kept hypoxic conditions and temperature constant for the duration of 18 days, representing a chronic exposure [ $> 4$  days; 16]. In our study population, at the onset of the reproductive season, individuals migrate into shallow protected bays with eelgrass, where algal overgrowth is common [77] so that the water may become hyperoxic during the day due to photosynthesis, but hypoxic at night due to algal and plant respiration [38-40, 57]. Thus, this population may experience hypoxic events that are high in frequency and magnitude but periodical in duration, conditions that are difficult to replicate in the laboratory. In the paternal egg-guarding Plainfin Midshipman fish, *Porichthys notatus*, a species that also reproduces in near-shore environments with fluctuating  $O_2$  levels, caring males are able to withstand hypoxia for twice as long as females [78]. Whether *S. typhle* males and females differ in their ability to tolerate hypoxic conditions, related to the sexual difference in parental care, is unknown but worth exploring in the future.

#### *Paternal care*

Brooding males provide nutrients, osmoregulation and oxygenation to the developing embryos. In addition, inside the brood pouch embryos are safe from external predation so that whole brood mortality depends on paternal survival, whereas individual embryo survival may depend on paternal brooding abilities (nutrients,  $O_2$  etc.) or embryo competition [45, 79]. In the current study, relative embryo survival was around 75% in both treatments; a number

comparable to previous studies on this population [48, 67], and importantly embryo survival did not differ between egg size or O<sub>2</sub> treatments. Does this mean males are able to adjust O<sub>2</sub> provision to the embryos in relation to embryonic consumption? Probably not significantly: while we do show that males brooding large eggs swim more than males brooding small eggs, which may facilitate paternal O<sub>2</sub> uptake and allow them to provide relatively more O<sub>2</sub> to larger embryos, we also know that pouch fluid O<sub>2</sub> is considerably lower than that of the tank water [68], both under hypoxia and normoxia. In addition, pouch O<sub>2</sub> drops markedly with time in the brooding period, that is, when O<sub>2</sub> consumption by the embryos is likely to be higher [68]. Therefore, if males are able to adjust O<sub>2</sub> provisioning to the embryos, such ability appears to be limited .

An alternative explanation for the lack of difference in embryo survival between broods consisting of the large and small egg size, and the lack of interaction between egg size and O<sub>2</sub> levels, is that with smaller egg size, more eggs fit in the pouches. Thus, if there is a benefit of being a small egg in terms of oxygenation, the larger number of embryos developing in close contact, which result in similar total egg surface areas in the two egg size treatments, may mask it. This could be tested in future studies, by keeping egg numbers rather than pouch fullness constant, while varying egg size. However, we chose not to do this in the current study, because if e.g. 100 large eggs fill a male's pouch, then 100 small eggs will leave it partially empty, this would have led to differences in pouch fluid volumes, and potentially also the amount of O<sub>2</sub> available in the fluid.

Why was embryo survival not affected by hypoxia in our study, when it has been documented in other aquatic species [e.g. 80, 81]? Our chosen level of 40% O<sub>2</sub> saturation (3.50 mg O<sub>2</sub>/L) may not have been low enough to affect embryo survival, even though it clearly affected

embryo size negatively and caused a significant increase in ventilation rates of brooding males. A range of O<sub>2</sub> concentrations have been used in other studies and species showing significant variation in embryo sensitivity to hypoxia [4, 81]. Yet, given that our chosen level of 40% is comparatively low for natural seagrass meadows, and the exposure in our experiment was considerably longer than what is common in nature, this result shows that *S. typhle* is able to cope with brooding also at reduced O<sub>2</sub> levels.

Broadly, our study raises the question as to whether pouch brooding in syngnathids evolved at least partly as an adaptation to hypoxia. It is common in fish for males that provide parental care during embryonic development to adjust their fanning behaviour in response to ambient O<sub>2</sub> conditions [33, 82] as well as to the developmental stage of the embryos [83]. In the seahorse, *Hippocampus zosterae*, males increase O<sub>2</sub> consumption during brooding by up to 50% compared to when not brooding [84]. Interestingly, in our study, day in the brooding period (i.e. embryonic stage) did not influence male ventilation rates or activity patterns. However, since embryonic metabolic demands tend to peak just before hatching [8, 46, 69, 85-87] it is possible that hypoxia impacts embryo survival only towards the end of the brooding period.

#### *The aquatic puzzle: egg size and oxygen under paternal care*

Pipefishes and seahorses display substantial variation among genera in where and how brood care is performed [37, 43]. Species with embryos attached to the surface of the male body (without a pouch) tend to have smaller eggs than species with enclosed pouches [37]. In the absence of a pouch embryos access O<sub>2</sub> directly from the ambient water, whereas in pouch brooders embryos depend on paternal oxygenation. In *S. typhle*, egg size is strongly correlated with female body length, with large females producing eggs that are substantially larger than

those of small females [37]. Despite this pronounced intraspecific variation in egg size and the knowledge that large embryos respire more than smaller ones [46], we found no support for the hypothesis that large eggs do worse than small eggs in hypoxia due to their lower surface area to volume ratio. We found that under hypoxia male ventilation rates increased significantly and that embryo length and weight were significantly and negatively affected. However, these effects were similar in both egg size classes and embryo survival at 18 days of age did not differ in relation to ambient O<sub>2</sub> levels. Thus, in contrast with long-established theory, embryos from large eggs were not more constrained in their development than embryos from smaller eggs in males brooding either in hypoxia or normoxia. This may be due to embryonic metabolic demands not increasing proportionally with egg volume, as discussed above, or to the pregnancy in *S. typhle*, where males are able to provide nutrients and O<sub>2</sub> and to osmoregulate the embryos during brooding. In the theoretical model developed by Jorgensen and colleagues (2012), ecological factors favouring the evolution of large offspring size also favour live bearing. In accordance with this, in *S. typhle* we find females producing large eggs for their body size [37] and males providing long and multifaceted paternal care in the safety of the brood pouch.

In conclusion, our study did not provide support for the surface-to-volume ratio argument for constraints in egg size evolution of aquatic organisms because, although *S. typhle* produces relatively large eggs and has limited ability to oxygenate the developing embryos, the observed negative effects of O<sub>2</sub> limitation on embryo development were largely independent of egg size. Thus, this hypothesised constraint is unlikely to be universal and the controversy around this issue is far from settled. Our study thus highlights the need for more intra- and interspecific empirical tests, on a greater variety of aquatic organisms, to better understand the evolutionary puzzle of egg size, oxygenation and parental care in aquatic environments.

466

467 **Author contributions:**

468 Conceived and designed the experiments: I.B.G., I.A. and C.K. Performed the experiments:

469 I.B.G. Analysed the data: I.B.G. Wrote the paper: I.B.G., I.A. and C.K.

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477

478 **Ethics statement:**

479 The experiment was done according to Swedish law, with an ethical approval given by the

480 Swedish Animal Welfare Agency (permits nr 196-2005 and 112-2007). This study does not

481 involve any endangered or protected species.

482

483 **Data accessibility:**

484 All data will be made accessible on Dryad Digital Repository.

485

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## LITERATURE CITED

1. Roff D. 1992 *The evolution of life histories: theory and analysis*. New York, Chapman & Hall.
2. Chambers R.C., Leggett W.C. 1996 Maternal influences on variation in egg sizes in temperate marine fishes. *Am Nat*, **36**, 180-196.
3. Smith C., Fretwell S. 1974 The optimal balance between size and number of offspring. *Am Nat*, **108**, 499-506.
4. Einum S., Hendry A.P., Fleming I.A. 2002 Egg-size evolution in aquatic environments: does oxygen availability constrain size? *Proc R Soc B*, **269**, 2325-2330. (doi:10.1098/rspb.2002.2150).
5. Hendry A.P., Day T. 2003 Revisiting the positive correlation between female size and egg size. *Evol Ecol Res*, **5**, 421-429.
6. Kolm N., Ahnesjö I. 2005 Do egg size and parental care coevolve in fishes? *J Fish Biol*, **66**, 1499-1515. (doi:10.1111/j.0022-1112.2005.00777.x).
7. Kranenbarg S., Verhagen J.H.G., Muller M., van Leeuwen J.L. 2001 Consequences of forced convection for the constraints on size and shape in embryos. *J Theor Biol*, **212**, 521-533.
8. Rombough P.J. 1988 Respiratory gas exchange, aerobic metabolism, and effects of hypoxia during early life. In *Fish physiology - The physiology of developing fish Part A Eggs and Larvae* (eds. Hoar W.S., Randall D.J.), pp. 59-161. London, Academic Press.
9. Naylor J.K., Taylor E.W., Bennett D.B. 1999 Oxygen uptake of developing eggs of *Cancer pagurus* (Crustacea: Decapod : Cancridae) and consequent behaviour of the ovigerous females. *J Mar Biol Assoc UK* **79**, 305-315.
10. Chaffee C., Strathmann R. 1984 Constraints on egg masses I. Retarded development within thick egg masses. *J Exp Mar Biol Ecol* **84**, 73-83.

11. Lee C.E., Strathmann R.R. 1998 Scaling of gelatinous clutches: effects of sibling competition for oxygen on clutch size and parental investment per offspring. *Am Nat*, **151**, 293-310. (doi:10.1086/286120 ).
12. Quinn T.P., Hendry A.P., Wetzel L.A. 1995 The influence of life history trade-offs and the size of incubation gravels on egg size variation in sockeye salmon (*Oncorhynchus nerka*). *Oikos*, **74**, 425-438.
13. Rombough P.J. 2007 Oxygen as a constraining factor in egg size evolution in salmonids. *Can J Fish Aquat Sci*, **64**, 692-699.
14. Pelster B. 1999 Environmental influences on the development of the cardiac system in fish and amphibians. *Comp Biochem Phys*, **124**, 407-412.
15. Reardon E.E., Chapman L.J. 2009 Hypoxia and life-history traits in a eurytopic African cichlid. *J Fish Biol*, **75**, 1795-1815. (doi:poll10.1111/j.1095-8649.2009.02429.x).
16. Pollock M.S., Clarke L.M.J., Dube M.G. 2007 The effects of hypoxia on fishes: from ecological relevance to physiological effects. *Environ Rev*, **15**, 1-14. (doi:10.1139/a06-006).
17. Bradford D.F., Seymour R.S. 1985 Energy conservation during the delayed hatching period in the frog *Pseudophryne bibroni*. *Physiol Zool*, **58**, 491-496.
18. Hamor T., Garside E.T. 1977 Size relations and yolk utilization in embryonated ova and alevins of Atlantic salmon *Salmo salar* L. in various combinations of temperature and dissolved oxygen. *Can J Zool*, **55**, 1892-1898.
19. Mills N.E., Barnhart M.C. 1999 Effects of hypoxia on embryonic development in two *Ambystoma* and two *Rana* species. *Physiol Biochem Zool* **72**, 179-188.
20. Cancino J.M., Gallardo J.A., Brante A. 2011 The relationship between temperature, oxygen condition and embryo encapsulation in the marine gastropod *Chorus giganteus*. *J Mar Biol Assoc UK*, **91**, 727-733. (doi:10.1017/s0025315410001335).



- 541        21.        Shine R. 1978 Propagule size and parental care: The "Safe Harbor" hypothesis *J*  
542        *Theor Biol*, **75**, 417-424.
- 543        22.        Sargent R.C., Taylor P.D., Gross M.R. 1987 Parental care and the evolution of  
544        egg size in fishes. *Am Nat*, **129**, 32-46.
- 545        23.        Summers K., McKeon C.S., Heying H. 2006 The evolution of parental care and  
546        egg size: a comparative analysis in frogs. **273**, 687-692. (doi:10.1098/rspb.2005.3368).
- 547        24.        Smith R.L. 1997 Evolution of paternal care in the giant water bugs (Heteroptera:  
548        Belostomatidae). In *The evolution of social behaviour in insects and arachnids* (eds. Crespi  
549        B.J., Choe J.C.), pp. 116-149. Cambridge, Cambridge University.
- 550        25.        Duarte C.M., Alcaraz M. 1989 To produce many small or few large eggs - a  
551        size-independent reproductive tactic of fish. **80**, 401-404.
- 552        26.        Jorgensen C., Auer S.K., Reznick D.N. 2011 A model for optimal offspring size  
553        in fish, including live-bearing and parental effects. *Am Nat*, **177**, E119-E135.  
554        (doi:10.1086/659622).
- 555        27.        Barlow G.W. 1981 Patterns of parental investment and size among coral-reef  
556        fishes. *Env Biol Fish*, **6**, 65-85. (doi:10.1007/BF00001801 ).
- 557        28.        Wisenden B.D., Unruii A., Morantes A., Bury S., Curry B., Driscoll R., Hussein  
558        M., Markegard S. 2009 Functional constraints on nest characteristics of pebble mounds of  
559        breeding male hornyhead chub *Nocomis biguttatus*. *J Fish Biol*, **75**, 1577-1585.  
560        (doi:10.1111/j.1095-8649.2009.02384.x).
- 561        29.        Takegaki T. 2001 Environmental factors affecting the spawning burrow  
562        selection by the gobiid *Valenciennea longipinnis*. *J Fish Biol*, **58**, 222-229.  
563        (doi:10.1006/jfbi.2000.1438).

- 564 30. Takegaki T., Nakazono A. 1999 Responses of the egg-tending gobiid fish  
565 *Valenciennea longipinnis* to the fluctuation of dissolved oxygen in the burrow. *Bull Mar Sci*,  
566 **65**, 815-823.
- 567 31. Jones J.C., Reynolds J.D. 1999 Costs of egg ventilation for male common  
568 gobies breeding in conditions of low dissolved oxygen. *Anim Behav*, **57**, 181-188.
- 569 32. Jones J.C., Reynolds J.D. 1999 Oxygen and the trade-off between egg  
570 ventilation and brood protection in the common goby. *Behaviour*, **136**, 819-832.
- 571 33. Lissåker M., Kvarnemo C. 2006 Ventilation or nest defense - parental care  
572 trade-offs in a fish with male care. *Behav Ecol Sociobiol*, **60**, 864-873. (doi:10.1007/s00265-  
573 006-0230-0).
- 574 34. Payne A.G., Smith C., Campbell A.C. 2004 A model of oxygen-mediated filial  
575 cannibalism in fishes. **174**, 253-266. (doi:10.1016/j.ecolmodel.2003.09.026).
- 576 35. Payne A.G., Smith C., Campbell A.C. 2002 Filial cannibalism improves survival  
577 and development of beaugregory damselfish embryos. *Proc R Soc B-Biol Sci*, **269**, 2095-  
578 2102. (doi:10.1098/rspb.2002.2144).
- 579 36. Braga Goncalves I. 2010 Egg size evolution and paternal care in pipefishes.  
580 Gothenburg, University of Gothenburg.
- 581 37. Braga Goncalves I., Ahnesjö I., Kvarnemo C. 2011 The relationship between  
582 female body size and egg size in pipefishes. *J Fish Biol* **78**, 1847-1854. (doi:10.1111/j.1095-  
583 8649.2011.02984.x ).
- 584 38. Greve T.M., Borum J., Pedersen O. 2003 Meristematic oxygen variability in  
585 eelgrass (*Zostera marina*). *Limnol Oceanogr*, **48**, 210-216.
- 586 39. Moore K.A. 2004 Influence of seagrasses on water quality in shallow regions of  
587 the lower Chesapeake Bay. *J Coast Res*, 162-178.

- 588        40.        Moore K.A., Jarvis J.C. 2008 Environmental factors affecting recent  
589 summertime eelgrass diebacks in the Lower Chesapeake Bay: Implications for long-term  
590 persistence. *J Coast Res*, 135-147. (doi:10.2112/si55-014).
- 591        41.        Cossellu M., Nordberg K. 2010 Recent environmental changes and filamentous  
592 algal mats in shallow bays on the Swedish west coast - A result of climate change? *J Sea Res*,  
593 **63**, 202-212. (doi:10.1016/j.seares.2010.01.004).
- 594        42.        Ripley J.L., Williams P.S., Foran C.M. 2010 Morphological and quantitative  
595 changes in paternal brood-pouch vasculature during embryonic development in two  
596 *Syngnathus* pipefishes. *J Fish Biol*, **77**, 67-79. (doi:10.1111/j.1095-8649.2010.02659.x).
- 597        43.        Stölting K.N., Wilson A.B. 2007 Male pregnancy in seahorses and pipefish:  
598 beyond the mammalian model. *Bioessays*, **29**, 884-896. (doi:10.1002/bies.20626).
- 599        44.        Carcupino M., Baldacci A., Mazzini M., Franzoi P. 2002 Functional  
600 significance of the male brood pouch in the reproductive strategies of pipefishes and  
601 seahorses: a morphological and ultrastructural comparative study on three anatomically  
602 different pouches. *J Fish Biol*, **61**, 1465-1480. (doi:10.1006/jfbi.2002.2160).
- 603        45.        Kvarnemo C., Mobley K.B., Partridge C., Jones A.G., Ahnesjö I. 2011 Evidence  
604 of paternal nutrient provisioning to embryos in broad-nosed pipefish *Syngnathus typhle*. *J*  
605 *Fish Biol*, **78**, 1725-1737. (doi:10.1111/j.1095-8649.2011.02989.x).
- 606        46.        Berglund A., Rosenqvist G., Svensson I. 1986 Reversed sex-roles and parental  
607 energy investment in zygotes of two pipefish (Syngnathidae) species. *Mar Ecol Prog Ser*, **29**,  
608 209-215. (doi:10.3354/meps029209).
- 609        47.        Ahnesjö I. 1992 Consequences of male brood care - weight and number of  
610 newborn in a sex-role reversed pipefish. *Func Ecol*, **6**, 274-281. (doi:10.2307/2389517).

48. Ahnesjö I. 1992 Fewer newborn result in superior juveniles in the paternally brooding pipefish *Syngnathus typhle* L. *J Fish Biol*, **41**, 53-63. (doi:10.1111/j.1095-8649.1992.tb03868.x).
49. Braga Goncalves I., Mobley K.B., Ahnesjö I., Sagebakken G., Jones A.G., Kvarnemo C. 2010 Reproductive compensation in broad-nosed pipefish females. *Proc R Soc B*, **277**, 1581-1587. (doi:10.1098/rspb.2009.2290).
50. Berglund A., Rosenqvist G., Svensson I. 1988 Multiple matings and paternal brood care in the pipefish *Syngnathus typhle*. *Oikos*, **51**, 184-188.
51. Jones A.G., Rosenqvist G., Berglund A., Avise J.C. 1999 The genetic mating system of a sex-role-reversed pipefish (*Syngnathus typhle*): a molecular inquiry. *Behav Ecol Sociobiol*, **46**, 357-365.
52. Jones A.G., Rosenqvist G., Berglund A., Avise J.C. 2000 Mate quality influences multiple maternity in the sex-role-reversed pipefish *Syngnathus typhle*. *Oikos*, **90**, 321-326.
53. Mobley K.B., Abou Chakra M., Jones A.G. 2014 No evidence for size-assortative mating in the wild despite mutual mate choice in sex-role-reversed pipefishes. *Ecol Evol*, **4**, 67-78. (doi:10.1002/ece3.907).
54. Sogabe A., Ahnesjö I. 2011 The ovarian structure and mode of egg production in two polygamous pipefishes: a link to mating pattern. *J Fish Biol*, **78**, 1833-1846. (doi:10.1111/j.1095-8649.2011.02973.x).
55. Berglund A., Rosenqvist G. 1990 Male limitation of female reproductive success in a pipefish - effects of body-size differences. *Behav Ecol Sociobiol*, **27**, 129-133. (doi:10.1007/BF00300646).
56. Ahnesjö I. 1995 Temperature affects male and female potential reproductive rates differently in the sex-role reversed pipefish, *Syngnathus typhle*. *Behav Ecol*, **6**, 229-233.

- 636        57.        Rheuban J.E., Berg P., McGlathery K.J. 2014 Multiple timescale processes drive  
637 ecosystem metabolism in eelgrass (*Zostera marina*) meadows. *Mar Ecol-Prog Ser*, **507**, 1-13.  
638 (doi:10.3354/meps10843).
- 639        58.        Lissåker M., Kvarnemo C., Svensson O. 2003 Effects of a low oxygen  
640 environment on parental effort and filial cannibalism in the male sand goby, *Pomatoschistus*  
641 *minutus*. *Behav Ecol*, **14**, 374-381.
- 642        59.        Anderson M.J., Gorley R.N., Clarke K.R. 2008 *PERMANOVA+ for PRIMER:*  
643 *Guide to software and statistical methods*. First edition ed. Plymouth, PRIMER-E Ltd.
- 644        60.        Clarke K.R., Gorley R.N. 2006 *PRIMER v6: User manual / Tutorial*. First  
645 edition ed. Plymouth, PRIMER-E Ltd.
- 646        61.        Berglund A., Rosenqvist G., Svensson I. 1986 Mate choice, fecundity and  
647 sexual dimorphism in two pipefish species (Syngnathidae). *Behav Ecol Sociobiol*, **19**, 301-  
648 307.
- 649        62.        Svensson I. 1988 Reproductive costs in two sex role reversed pipefish species  
650 (Syngnathidae). *J Anim Ecol*, **57**, 929-942.
- 651        63.        Kvarnemo C., Moore G.I., Jones A.G. 2007 Sexually selected females in the  
652 monogamous Western Australian seahorse. *Proc R Soc B*, **274**, 521-525.  
653 (doi:10.1098/rspb.2006.3753).
- 654        64.        Parsons G.R., Carlson J.K. 1998 Physiological and behavioral responses to  
655 hypoxia in the bonnethead shark, *Sphyrna tiburo*: routine swimming and respiratory  
656 regulation. *Fish Physiol Biochem*, **19**, 189-196. (doi:10.1023/a:1007730308184).
- 657        65.        Roberts J.L. 1975 Active branchial and ram gill ventilation in fishes *Biol Bull*,  
658 **148**, 85-105. (doi:10.2307/1540652).

- 659        66.        Munguia-Steyer R., Favila M.E., Macias-Ordonez R. 2008 Brood pumping  
660 modulation and the benefits of paternal care in *Abedus breviceps* (Hemiptera :  
661 Belostomatidae). *Ethology*, **114**, 693-700. (doi:10.1111/j.1439-0310.2008.01507.x).
- 662        67.        Ahnesjö I. 1996 Apparent resource competition among embryos in the brood  
663 pouch of a male pipefish. *Behav Ecol Sociobiol*, **38**, 167-172.
- 664        68.        Braga Goncalves I., Ahnesjö I., Kvarnemo C. 2015 Embryo oxygenation in  
665 pipefish brood pouches: novel insights. *J Exp Biol*, **218**, 1639-1646.
- 666        69.        Hamor T., Garside E.T. 1979 Hourly and total oxygen consumption by ova of  
667 Atlantic salmon, *Salmo salar* L., during embryogenesis, at two temperatures and three levels  
668 of dissolved oxygen. *Can J Zool*, **57**, 1196-1200. (doi:10.1139/z79-152 ).
- 669        70.        Fernández M., Pappalardo P., Jeno K. 2006 The effects of temperature and  
670 oxygen availability on intracapsular development of *Acanthina monodon* (Gastropoda:  
671 Muricidae). *Rev Chil Hist Nat* **79**, 155-167.
- 672        71.        Kolding J., Haug L., Stefansson S. 2008 Effect of ambient oxygen on growth  
673 and reproduction in Nile tilapia (*Oreochromis niloticus*). *Can J Fish Aquat Sci* **65**, 1413-1424.  
674 (doi:10.1139/f08-059).
- 675        72.        Kamler E. 2008 Resource allocation in yolk-feeding fish. *Rev Fish Biol Fish*,  
676 **18**, 143-200. (doi:10.1007/s11160-007-9070-x).
- 677        73.        Richmond C., Marcus N.H., Sedlacek C., Miller G.A., Oppert C. 2006 Hypoxia  
678 and seasonal temperature: short-term effects and long-term implications for *Acartia tonsa*  
679 *dana*. *J Exp Mar Biol Ecol*, **328**, 177-196.
- 680        74.        Breitburg D.L., Hondorp D.W., Davias L.A., Diaz R.J. 2009 Hypoxia, nitrogen,  
681 and fisheries: integrating effects across local and global landscapes. *Annu Rev Mar Sci*, **1**, 329-  
682 349. (doi:10.1146/annurev.marine.010908.163754).

- 683        75.        Doney S.C., Ruckelshaus M., Duffy J.E., Barry J.P., Chan F., English C.A.,  
684        Galindo H.M., Grebmeier J.M., Hollowed A.B., Knowlton N., et al. 2012 Climate Change  
685        Impacts on Marine Ecosystems. *Annu Rev Mar Sci*, **4**, 11-37. (doi:10.1146/annurev-marine-  
686        041911-111611).
- 687        76.        Poertner H.O., Peck M.A. 2010 Climate change effects on fishes and fisheries:  
688        towards a cause-and-effect understanding. *J Fish Biol*, **77**, 1745-1779.
- 689        77.        Pihl L., Svensson A., Moksnes P.-O., Wennhage H. 1999 Distribution of green  
690        algal mats throughout shallow soft bottoms of the Swedish Skagerrak archipelago in relation  
691        to nutrient sources and wave exposure. *J Sea Res*, **41**, 281-294.
- 692        78.        LeMoine C.M.R., Bucking C., Craig P.M., Walsh P.J. 2014 Divergent hypoxia  
693        tolerance in adult males and females of the plainfin midshipman (*Porichthys notatus*). *Physiol*  
694        *Biochem Zool*, **87**, 325-333. (doi:10.1086/674565).
- 695        79.        Sagebakken G., Ahnesjö I., Mobley K.B., Goncalves I.B., Kvarnemo C. 2010  
696        Brooding fathers, not siblings, take up nutrients from embryos. *Proc R Soc B*, **277**, 971-977.  
697        (doi:10.1098/rspb.2009.1767).
- 698        80.        Keckeis H., BauerNemeschkal E., Kamler E. 1996 Effects of reduced oxygen  
699        level on the mortality and hatching rate of *Chondrostoma nasus* embryos. *J Fish Biol*, **49**,  
700        430-440.
- 701        81.        Diez J.M., Davenport J. 1990 Energy exchange between the yolk and embryo of  
702        dogfish (*Scyliorhinus canicula* L.) eggs held under normoxic, hypoxic and transient anoxic  
703        conditions *Comp Biochem Physiol* **96**, 825-830.
- 704        82.        Hale R.E., St Mary C.M., Lindstrom K. 2003 Parental responses to changes in  
705        costs and benefits along an environmental gradient. *Environ Biol Fish*, **67**, 107-116.

83. Green B.S., McCormick M.I. 2005 O<sub>2</sub> replenishment to fish nests: males adjust brood care to ambient conditions and brood development. *Behav Ecol*, **16**, 389-397. (doi:10.1093/beheco/ari007).

84. Masonjones H.D. 2001 The effect of social context and reproductive status on the metabolic rates of dwarf seahorses (*Hippocampus zosterae*). *Comp Biochem Physiol A-Mol Integr Physiol*, **129**, 541-555.

85. Cronin E.R., Seymour R.S. 2000 Respiration of the eggs of the giant cuttlefish *Sepia apama*. *Mar Biol*, **136**, 863-870.

86. Davenport J. 1983 Oxygen and the developing eggs and larva of the lumpfish, *Cyclopterus lumpus*. *J Mar Biol Assoc UK* **63**, 633-640. (doi:10.1017/S0025315400070946).

87. Diez J.M., Davenport J. 1987 Embryonic respiration in the dogfish (*Scyliorhinus canicula* L). *J Mar Biol Assoc UK*, **67**, 249-261.



Table 1 – Permutational MANOVA and ANOVA analyses of the effects of year (2007, 2008), egg size treatment (small vs. large) and oxygen treatment (100% vs. 40%) on *Syngnathus typhle*, a) overall and b) separately for relative embryo survival, average embryo length (mm) and dry weight (mg). N = 46. Analyses were performed on transformed and normalised variables.

**a) Multivariate test:**

Source	df	MS	Pseudo-F	p (perm)
Year	1	8.92	3.80	0.018 <sup>y</sup>
Egg size	1	10.16	4.32	0.011 <sup>y</sup>
Oxygen level	1	16.64	7.08	0.001 <sup>y</sup>
Egg size x Oxygen level	1	0.07	0.03	0.989 <sup>y</sup>
Residual	41	2.35 <sup>x</sup>		

**b) Univariate tests:**

Source	df	Survival			Length			Weight		
		MS	Pseudo-F	p (perm)	MS	Pseudo-F	p (perm)	MS	Pseudo-F	p (perm)
Year	1	3.89	4.12	0.049 <sup>y</sup>	5.01	7.03	0.011 <sup>y</sup>	0.01	0.02	0.892 <sup>y</sup>
Egg size	1	0.32	0.34	0.565 <sup>y</sup>	2.23	3.13	0.081 <sup>y</sup>	7.61	11.00	0.002 <sup>y</sup>
Oxygen level	1	0.26	0.28	0.595 <sup>y</sup>	8.40	11.78	0.001 <sup>y</sup>	7.98	11.54	0.002 <sup>y</sup>
Egg size x Oxygen level	1	0.06	0.07	0.802 <sup>y</sup>	0.00	0.00	0.997 <sup>y</sup>	0.01	0.01	0.920 <sup>y</sup>
Residual	41	0.95 <sup>x</sup>			0.71 <sup>x</sup>			0.69 <sup>x</sup>		

<sup>y</sup> - Term mean squares tested against the pooled mean squares of residuals and remaining non-significant interaction terms.

<sup>x</sup> – Mean square of pooled residuals and remaining non-significant interaction terms.

Table 2 – Permutational MANOVA and ANOVA analyses of the effects of egg size treatment (small vs. large), oxygen treatment (100% vs. 40%), year (2007, 2008), day (1, 9 and 18) and male ID (nested within egg size, oxygen level and year) on *Syngnathus. typhle* a) overall and b) separately for ventilation rates, proportion of time spent swimming and proportion of time spent in the upper half of the aquarium. N = 41.

a) Multivariate test:

Source	df	MS	Pseudo-F	p (perm)
Egg size	1	9.29	3.14	0.037 <sup>y</sup>
Oxygen level	1	44.21	14.94	< 0.001 <sup>y</sup>
Year	1	7.87	2.66	0.054 <sup>y</sup>
Day	2	14.61	2.71	0.177
Year x Day	2	5.38	2.68	0.032 <sup>y</sup>
Male	36	2.94	1.47	0.019 <sup>y</sup>
Residual	79	2.01 <sup>x</sup>		

b) Univariate tests:

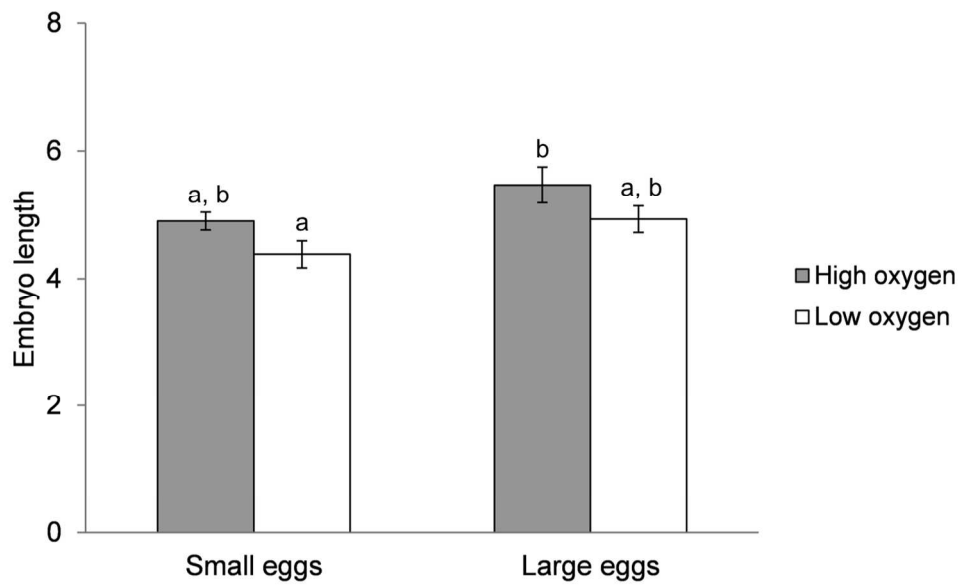
Source	Ventilation rate				Proportion of time swimming				Proportion of time in upper part of tank			
	df	MS	Pseudo-F	p (perm)	df	MS	Pseudo-F	p (perm)	df	MS	Pseudo-F	p (perm)
Egg size	1	0.00	0.00	0.977 <sup>y</sup>	1	7.08	7.49	0.010 <sup>y</sup>	1	2.21	2.43	0.132 <sup>y</sup>
Oxygen level	1	43.45	39.39	< 0.001 <sup>y</sup>	1	0.23	0.24	0.631 <sup>y</sup>	1	0.52	0.57	0.450 <sup>y</sup>
Year	1	1.96	1.78	0.201 <sup>y</sup>	1	5.87	6.21	0.017 <sup>y</sup>	1	0.04	0.04	0.843 <sup>y</sup>
Day	2	4.41	13.61	0.105	2	0.43	0.25	0.851	2	9.77	2.91	0.239
Year x Day	2	0.32	1.13	0.324 <sup>y</sup>	2	1.70	1.84	0.164 <sup>y</sup>	2	3.36	4.21	0.021 <sup>y</sup>
Male	36	1.10	3.85	< 0.001 <sup>y</sup>	36	0.95	1.02	0.452 <sup>y</sup>	36	0.91	1.14	0.317 <sup>y</sup>
Residual	79	0.29 <sup>x</sup>			79	0.93 <sup>x</sup>			79	0.80 <sup>x</sup>		

<sup>y</sup> - Term mean square was tested against pooled mean square of residuals and all other interactions.

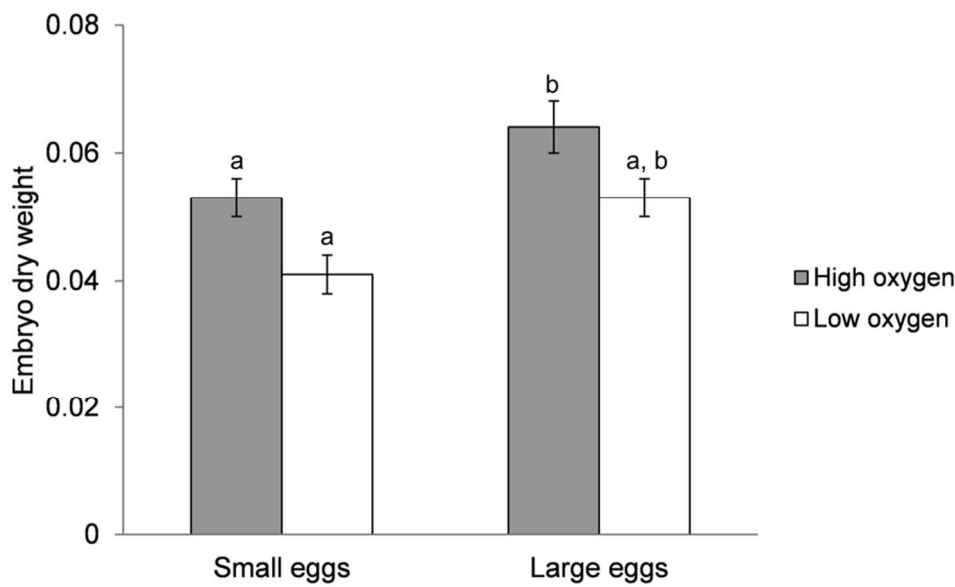
<sup>x</sup> – Mean square of pooled residuals and non-significant interaction terms.

Figure 1: a) Average embryo length (mm) and b) average embryo dry weight (mg) of *Syngnathus typhle* males that received small or large eggs and were kept either in high (100%) or low (40%) oxygen conditions for a brooding period of 18 days. Significant differences are displayed with different letters, based on LSD post-hoc tests following 1-way ANOVA, using an oxygen level-egg size composite treatment factor with four levels.

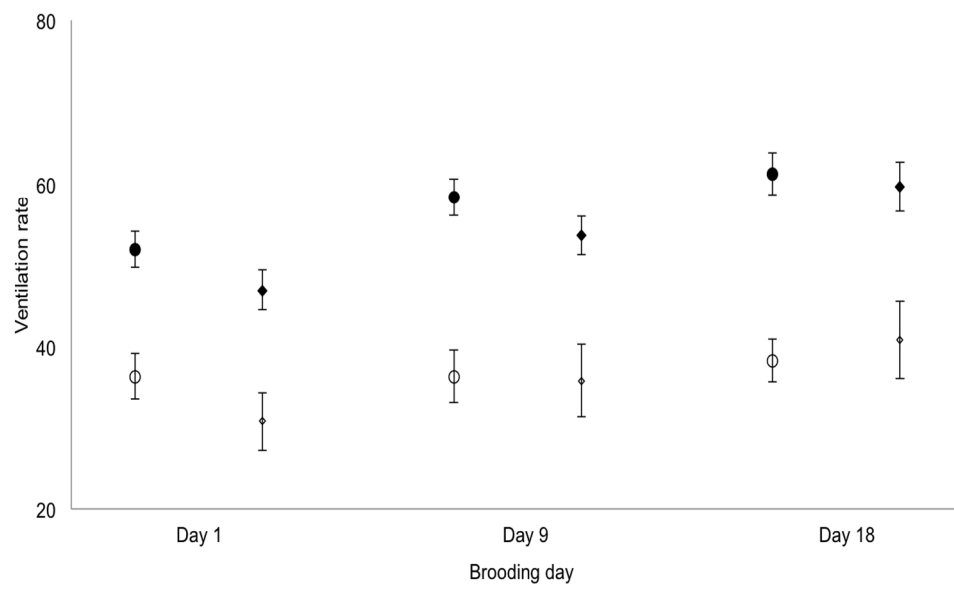
Figure 2: Average (mean  $\pm$  SE, ventilations per minute) ventilation rates of *Syngnathus typhle* males brooding embryos from large (circles) or small (diamonds) eggs, kept in high (open symbols) or low (filled symbols) oxygen conditions. Ventilation rates were recorded three times during the experiment, on days 1, 9 and 18.



952x571mm (96 x 96 DPI)



952x571mm (96 x 96 DPI)



960x576mm (96 x 96 DPI)